



Are you filling your chambers to capacity?

You may be starving your plants.

By Patrick Friesen, PhD

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You may be starving your plants. Why it matters to consider fresh air flow.

Maintaining CO₂ concentrations in the plant canopy can be critical for normal growth and development. Plants photosynthesize and consume CO₂ during growth. In contained environments such as plant growth chambers, the consumed CO₂ must be replenished in order to prevent appreciable drops in CO₂ concentrations. Appreciable drops in CO₂ concentrations can lead to reductions in biomass, delay or prevent flowering, and alter gene expression. Introducing ambient air into plant growth chambers is one method to replenish the CO₂.

How do plant area and fresh air flow rates affect CO₂ concentrations?

The more plant area inside a growth chamber, the greater the depletion of CO₂ (Fig. 1). The rate of CO₂ depletion is related to the total net CO₂ assimilation rate (rate of photosynthesis) of the entire plant area. With well-watered plants, light intensity, temperature, and the CO₂ concentration inside the chamber itself primarily affect the rate of CO₂ assimilation¹.

If growth chambers are filled to capacity, the fresh air flow rates through the chambers have a considerable effect on the CO₂ concentrations inside. With their standard fresh air flow rates, CO₂ concentrations dropped to 280ppm inside both a reach-in chamber filled with field mustard (*Brassica rapa*)² and a walk-in growth room filled with poplar trees (*Populus* sp.)³. In chambers built before 1984, fresh air flow rates are generally even less and can result in CO₂ concentrations of 150ppm and 50ppm inside a chamber filled with cotton (*Gossypium* sp.) and maize (*Zea mays*) respectively³⁻⁵. Increased fresh air flow rates increase the CO₂ concentration around plants by replacing the CO₂ consumed during photosynthesis at a greater rate (Fig. 2).

A BioChambers short plant chamber (model SPC-37) filled with young maize and soybean plants provides a fresh air flow rate that exceeds 50 ft³ min⁻¹, mitigating CO₂ depletion to less than 40ppm (Fig. 3). Assuming an ambient CO₂ concentration close to atmospheric (~400ppm), the 51 ft³ min⁻¹ fresh air flow rate of the SPC-37 is within this 10% CO₂ drawdown, the most stringent recommendation of Morse (1963)⁶ in the **Plant Growth Chamber Handbook**⁷.

In addition, we estimate net CO₂ assimilation rates per unit growth area for plants that use C₃ and C₄ photosynthesis at various growth temperatures using leaf level gas exchange data and comparable whole chamber experiments for a tall plant chamber (model TPC-19). These estimates were used to calculate the required fresh air flow rates to achieve a 10% CO₂ drawdown and are based on an assumed leaf area index (leaf area/growth area, m² m⁻²) of 1.5 for tall plants (Table 1). Theoretical estimates for the SPC-37 using this same approach are ±1.3 ft³ min⁻¹ of actual measurements from the experiment with maize and soybean.

How do low CO₂ concentrations and inadequate fresh air intake affect plant growth and development?

When CO₂ concentrations inside growth chambers drop below ambient (~400ppm), growth and development for plants that use C₃ photosynthesis are reduced and impaired (eg. wheat, soybean, rice)¹⁶⁻²⁶. The majority of plant species use the C₃ photosynthetic pathway, where at low CO₂ concentrations, CO₂ itself becomes the primary limitation on carbon gain. Ribulose-1-5 carboxylase/oxygenase (Rubisco) is the photosynthetic enzyme that fixes CO₂ into sugars for growth and development. When CO₂ concentrations are low, Rubisco and carbon gain are limited by CO₂^{27, 1}. Rubisco also reacts with O₂ in the energetically wasteful process of photorespiration²⁸. Here O₂ competes with and inhibits CO₂ binding, exacerbating the CO₂ limitation on carbon gain as CO₂ concentrations decline, especially at warm temperatures²⁹⁻³². Across a number of plants that use C₃ photosynthesis, reductions in biomass are roughly proportional to reductions in CO₂ concentrations below current atmospheric levels (~400ppm). Here, a 50% reduction in CO₂ concentration results in a 50% reduction in biomass (see ref. 24, Fig. 3). Other effects of low CO₂ concentrations on C₃ plants include increased water use, a reduction in the ratio of root:shoot biomass, and delayed or failure to flower (reviewed in ref. 19). The stress of low CO₂ concentration affects gene expression and quantitative traits, potentially confounding the expression profiles from treatment factor(s) of interest². Plants that can concentrate CO₂ around Rubisco experience reduced effects of low CO₂ concentrations on growth and development, such as plants that use C₄ photosynthesis (eg. maize, sugarcane)^{18, 23}.

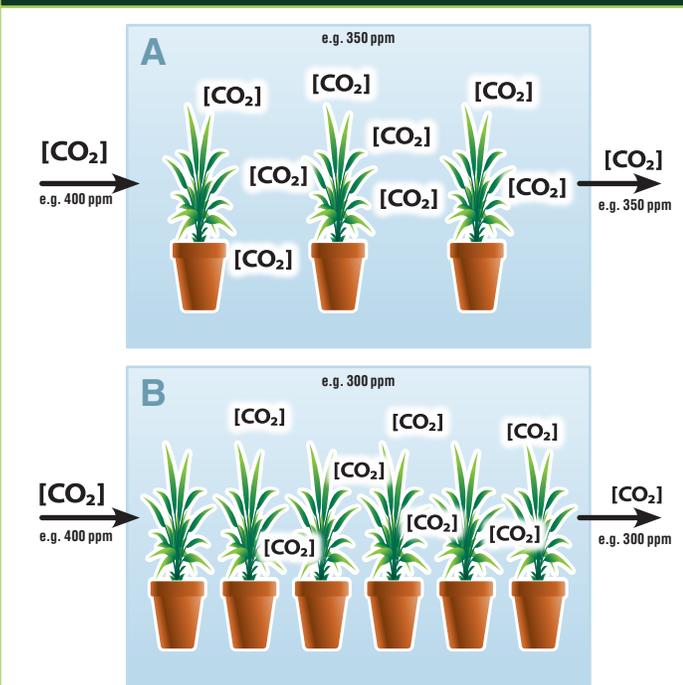
In addition, gases emitted by plants such as isoprene and ethylene can potentially build up and have feedback effects if fresh air flow is inadequate. These feedbacks could have undesirable effects and further confound experiments. Increased isoprene exposure can hasten the onset of flowering and may artificially increase tolerance of high temperatures³³⁻³⁵. Low CO₂ environments can increase isoprene emissions, further compounding the problem of inadequate fresh air supply³⁶. Superambient ethylene exposure accelerates fruit ripening and can slow or accelerate plant growth depending on the species^{37, 38}.

Photosynthetic Type	Leaf Temp (°C)	Required Fresh Air Flow Rate (ft ³ min ⁻¹)
C ₃ (eg. wheat)	21	31
C ₃ (eg. wheat)	30	33
C ₃ (eg. wheat)	35	29
C ₄ (eg. maize)	21	28
C ₄ (eg. maize)	30	44
C ₄ (eg. maize)	35	46

Table 1:

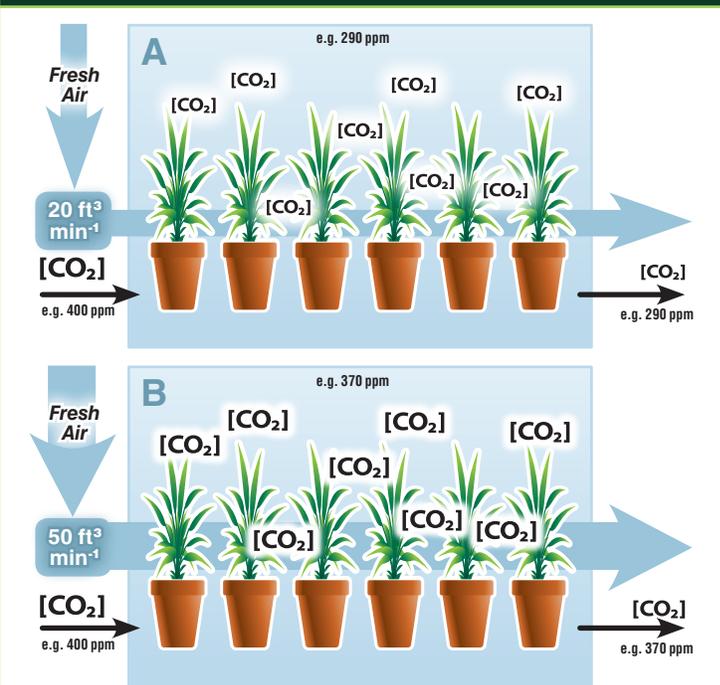
Calculated fresh air flow rates (ft³ min⁻¹) required to maintain a 10% reduction in ambient CO₂ concentration inside a tall plant chamber (model TPC-19) filled to capacity for plants that use C₃ or C₄ photosynthesis at different growth temperatures. Calculations assume a 400ppm CO₂ concentration entering the chamber and a leaf area index (leaf area/growth area, m² m⁻²) of 1.5. Estimates are based on published leaf level net CO₂ assimilation rates and comparable whole chamber CO₂ assimilation studies⁸⁻¹⁵. These estimates incorporate the maximum photosynthetic photon flux density and growth area of the TPC-19 which are 1500 μmol m⁻² s⁻¹ and 1.87m² respectively. The TPC-19 has a fresh air flow rate of 60 ft³ min⁻¹, exceeding these requirements under the given assumptions.

Figure 1



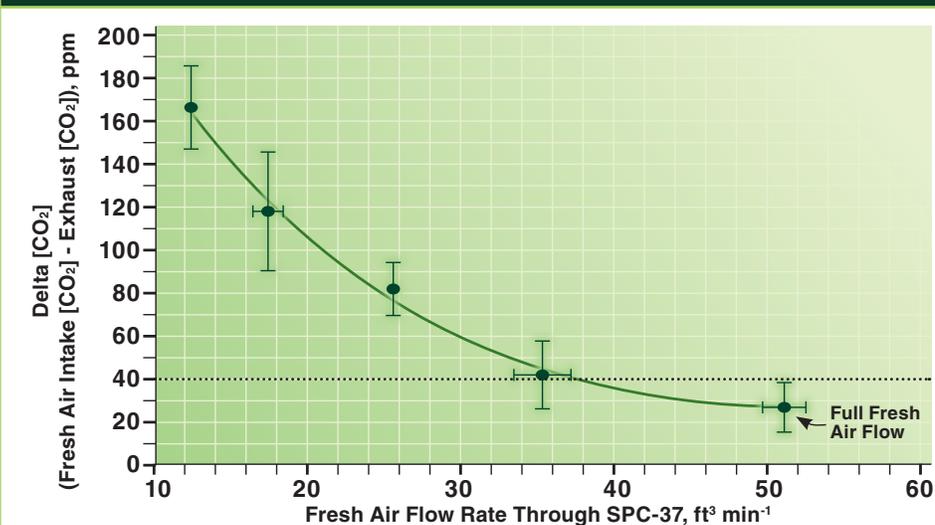
The effect of greater plant mass/area on CO₂ concentration inside a growth chamber. During photosynthesis plants consume CO₂ from the surrounding air. The greater the leaf area inside a growth chamber, the greater the rate of CO₂ drawdown from photosynthesis. Six plants (B) will lower the CO₂ concentration more than only three plants (A) of equivalent size and development.

Figure 2



The effect of greater fresh air flow on the CO₂ concentration inside a growth chamber. Greater fresh air flow replaces the CO₂ consumed during photosynthesis at a faster rate, increasing the CO₂ concentration or mitigating the drawdown from photosynthesis. With an equivalent number of plants of the same size and development, increasing the fresh air flow from 20 ft³ min⁻¹ (A), to 50 ft³ min⁻¹ (B), can prevent a nearly four-fold internal drawdown of CO₂ concentrations (see Figure 3).

Figure 3



Drawdown of CO₂ concentration as a function of fresh air flow inside a BioChambers SPC-37 filled with well-watered and fertilized maize and soybean (mean ±SE). Leaf temperatures ranged from 25.5 – 26.5°C and photosynthetic photon flux densities averaged 430 μmol m⁻² s⁻¹ across the upper leaves. The leaf area index (leaf area/growth area, m² m⁻²) of all plants was 0.48. Flow rates were decreased by manually closing the fresh air intake valve from fully open (arrow). After each flow rate change, at least 45 minutes was given before measurements were recorded to allow for steady state conditions. Dotted line is the 10% recommended drawdown limit of Morse (1963)⁶ assuming the ambient CO₂ concentration entering the chamber is current atmospheric (~400ppm).

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